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Aryl- and heteroaryl-substituted aminobenzo[a]quinolizines as dipeptidyl peptidase IV inhibitors

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ABSTRACT

Synthesis and SAR are described for a structurally distinct class of DPP–IV inhibitors based on aminobenzo[a]quinolizines bearing (hetero-)aromatic substituents in the S1 specificity pocket. The m-(fluoromethyl)-phenyl derivative (S,S,S)-2g possesses the best fit in the S1 pocket. However, (S,S,S)-2i, bearing a more hydrophilic 5-methyl-pyridin-2-yl residue as substituent for the S1 pocket, displays excellent in vivo activity and superior drug-like properties.

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Dipeptidyl peptidase IV (DPP–IV) inhibitors have emerged as a new therapeutic option to treat patients with type 2 diabetes. ^{1,2} The pharmacological action is based mainly on the ability to prevent cleavage and inactivation of glucagon-like peptide (GLP–1), an incretin hormone which stimulates insulin secretion and inhibits glucagon secretion.

Several DPP–IV inhibitors have advanced into late stage clinical studies.^{2–4} Of these, sitagliptin⁵ (**1a**, Fig. 1), vildagliptin⁶ and saxagliptin⁷ have been approved and launched in several countries. While DPP–IV inhibitors encompass a remarkable structural diversity, with a primary or secondary amino group being the sole recurring motif, they all possess substituents which are able to occupy the S1 specificity pocket of the target. For instance, the trifluorophenyl subunit of **1a** fully occupies the S1 pocket of DPP–IV.

We have recently reported the discovery of aryl-substituted aminobenzo[a]quinolizines **2a-c** (Fig. 1), a structurally distinct class of DPP-IV inhibitors. These inhibitors were derived from high-throughput screening hit **3**, a legacy compound with favourable molecular properties, which was synthesised in the 1950s as one member of a series of emetine analogues. X-ray structural information was used together with structure-activity relationships obtained for simplified 1,3-substituted 4-aminopiperidines

and aminomethyl-pyrimidines⁹ such as **4a** as the key elements in the lead identification process culminating in **2b** and **2c**, the first highly potent representatives of this class. Like **1a** and **4a**, **2b** and **2c** possess a substituted benzene ring as ligand for the S1 pocket. Here, we report the further evolution of the class culminating in (*S*,*S*,*S*)-**2i**, a highly potent compound with a more balanced profile.

Having established that a lipophilic substituent at the *meta*-position of the aromatic ring (R¹ in compound **2**) brings about a significant improvement in activity by occupying the lipophilic niche

Figure 1. Selected DPP-IV inhibitors.

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at the back of the S1 specificity pocket, we sought to explore alternative substituents on the aromatic ring. Also, there is room at the aromatic position para to R^1 (substituent X in compound **2**, Scheme 1) that could be occupied, either by a lipophilic substituent or by a heteroatom.

Therefore, we synthesised a set of aryl- and heteroaryl derivatives of the general formula **2** according to Scheme 1. Thus, tricyclic ketone **5**¹¹ was regio- and diastereoselectively arylated with halides **6** under palladium catalysis, affording only the 3 β , 11 β -stereoisomer **7**. Conversion of the keto group of **7** to the oxime followed by reduction produced mixtures of the 2 α and 2 β amine epimers in a ratio of ca 8:1. The 2 α -aminobenzo[α]quinolizines **2a**- β were separated from the less active 2 β -epimers by column chromatography.

The activities of **2a-i** as DPP-IV inhibitors are summarised in Table 1. As anticipated from the rigid nature of the S1 specificity pocket, the structure-activity relationship at R¹ is rather steep. A chlorine atom or methyl group improve the activity by 40-50-fold (2b, c vs 2a).8 However, the advantage over the unsubstituted phenyl residue in 2a is partly lost when R1 is a slightly larger alkyl group [ethyl (2d), cyclopropyl (2c)]. Even larger R¹ groups, such as isopropyl (2f), are not tolerated at all. On the other hand, one order of magnitude improvement in activity over 2c is gained by addition of a single fluorine atom (2g). Introduction of a methyl group at position X leads to a slight improvement in activity (ca two-fold for 2,5-dimethylphenyl derivative 2h). This effect is significant but smaller than that observed for electronegative groups like the ortho-chlorine in **4a**⁹ (140-fold improvement over **4b**) or for the ortho-fluorine in 1a (seven-fold improvement over 1b)5 which benefit from a favourable electrostatic interaction with the positively charged side chain of Arg125. 13 The polar environment afforded by the side chains of Arg125 and Asn710 also tolerates the incorporation of an ortho-pyridine. In fact, the pyridine derivatives 2i and 2i are only slightly less active than the benzene counterparts 2c and 2g although they are around one order of magnitude less lipophilic.

We were able to solve the X-ray complex structure of the fluoromethyl-substituted inhibitor ${\bf 2g}$ bound to human DPP-IV to a resolution of 2.8 Å (Fig. 2). The binding mode of the aminobenzo[a]quinolizine subunit is nearly identical to that of the HTS hit ${\bf 1},^{13,16}$ in that the aromatic ring makes a favourable, parallel-shifted $\pi-\pi$ interaction with Phe357, and the positively charged amino group is engaged in a tight H-bonding network with Glu205, Glu206, and Tyr662. The hydrophobic S1 pocket is fully occupied by the (fluoromethyl)-benzene moiety of ${\bf 2g}$. In particular, the CH₂F group nicely fits the asymmetry of the S1 pocket, with five van der Waals contacts to the side chains of Tyr631, Val656, Trp659, and Tyr666. The van der Waals interaction strength for the fluorine is about twice that of the hydrogen, thus rationalising the affinity improvement of ${\bf 2g}$ in comparison with ${\bf 2c}$.

Compound **2c** is a highly potent compound with a favourable set of drug-like properties. It is highly soluble and permeable, metabolically stable, and has only a minimal potential for drug-drug interaction. In vivo, it is active at an oral dose of 0.3 mg kg $^{-1}$ in the oral glucose tolerance test (OGTT) in Zucker fatty (fa/fa) rats, reducing glucose levels by 41% at a time-point of 40 min after the glucose challenge. On the downside, in vivo clearance and vol-

ume of distribution are very high (Table 2), and brain uptake is extensive, which is not a desirable property for a compound

Table 1
Structure–activity relationship of racemic compounds 2

Compd	Х	R ¹	IC_{50}^{a} (nM)	$\log D_{7.4}^{\mathrm{b}}$	in vitro PL ^c
2a	CH	Н	200 ^d	0.8	5
2b	CH	Cl	4.2 ^d	1.7	n.t.
2c	CH	CH ₃	4.6 ^d	1.3	2.5
2d	CH	CH ₂ CH ₃	61	1.7	n.t.
2e	CH	Cyclopropyl	38	1.8	n.t.
2f	CH	Isopropyl	5500	2.0	n.t.
2g	CH	CH ₂ F	0.5	1.6	n.t.
2h	C-CH ₃	CH ₃	1.7	1.6	n.t.
2i	N	CH ₃	19	0.3	15
2j	N	CH ₂ F	1.2	0.1	n.t.

^a For a description of experimental details for the determination of DPP-IV inhibition see: Ref. 8.

d Data from Ref. 8.

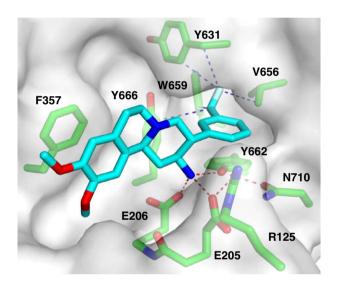


Figure 2. X-ray complex crystal structure of human DPP-IV with compound (S,S,S)-**2g.** Hydrogen bonds are displayed in red and van der Waals interactions $(d < \sum r_v dW + 0.5 \text{ Å})$ of the CH₂F moiety with DPP-IV are highlighted in blue.¹⁷

Table 2Pharmacokinetic parameters (Wistar rat) for selected compounds

Compd	Cl ^{a,b}	$V_{ss}^{a,c}$	$t_{1/2}^{\mathrm{a,d}}$	F ^e	B/P ^f
2c	87	42	6.9	56	5.8
(S,S,S)-2i	118	12	1.4	50	0.7

a Dose: 1 mg kg⁻¹ i.v.

^b Clearance [mL min⁻¹ kg⁻¹].

c Volume of distribution at steady state, [L kg⁻¹].

d Half life [h].

e Oral bioavailability [%] at 3 mg kg⁻¹.

f Brain to plasma ratio.

Scheme 1. Reagents and conditions: (a) Pd(OAc)₂, P'Bu₃, NaO'Bu, THF, 21-51%; (b) NH₂OH'HCl, NaOAc, EtOH, 83-99%; (c) H₂, Raney Ni, aq NH₃, MeOH, THF, 34-85%.

b Distribution coefficient at pH 7.4.14

 $[^]c$ Lowest concentration [μM] to induce phospholipidosis in vitro (test range 2.5–20 $\mu M).^{15}$

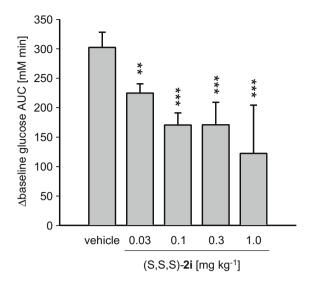


Figure 3. Dose-dependent effect of (S,S,S)-**2i** on glucose levels (area under the curve, $t = 0 \rightarrow 120$ min) during an oral glucose tolerance test in fa/fa rats. The compound was applied 2 h before the glucose challenge (2 g kg $^{-1}$). ** $p \le 0.01$, *** $p \le 0.001$, ANOVA followed by Dunnett's PostHoc test versus vehicle.

intended for a peripheral mode-of-action. Finally, 2c strongly induces phospholipidosis in cultured fibroblasts¹⁵ at the lowest tested concentration of 2.5 μ M.

A proven strategy to reduce the risk of phospholipidosis is to minimise the free energy of amphiphilicity, $\Delta\Delta G_{am}.^{18,19}$ Indeed, the pyridine analogue 2i ($\Delta\Delta G_{am}$ = -6.02 kJ mol $^{-1}$) is significantly less amphiphilic than 2c ($\Delta\Delta G_{am}$ = -6.41 kJ mol $^{-1}$). Accordingly, the lowest concentration of 2i to induce phospholipidosis in vitro is 15 μ M. Fluorine substitution on the pyridine methyl of 2i enhances the potency further (2j), but this compound unfortunately tested positive in the Ames test for mutagenicity 20 and in the micronucleus test for clastogenicity 21 and was thus not considered further.

The racemate **2i** was separated into its enantiomers using chiral HPLC (Chiralpak® AD). The (S,S,S)-enantiomer is potent ($IC_{50} = 11 \text{ nM}$), whereas the (R,R,R)-enantiomer is virtually inactive ($IC_{50} = 53,000 \text{ nM}$).

In comparison with lead compound 2c, (S,S,S)-2i exhibits superior pharmacokinetic properties, having a lower volume of distribution, less brain penetration, and a shorter half life (Table 2). In vivo, (S,S,S)-2i reduces glucose levels during an oral glucose tolerance test (OGTT) in Zucker fatty (fa/fa) rats in a dose-dependent manner (Fig. 3). Indeed, the compound already shows appreciable glucose reduction at a dose as low as 0.03 mg kg^{-1} .

In summary, the X-ray structure of DPP–IV suggested that potency and molecular properties in the aryl-aminobenzo[a]quinolizine series can be modulated through optimisation of the S1 substituent. Compound (*S*,*S*,*S*)-**2g**, bearing a *meta*-(fluoromethyl)-phenyl subunit, has the best fit in the S1 pocket. However, improved drug-like properties were obtained by installing a more polar pyridine ring in the S1 ligand position, leading to (*S*,*S*,*S*)-**2i**, a highly potent DPP–IV inhibitor with more balanced properties.

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References and notes

- 1. Drucker, D. J.; Nauck, M. A. Lancet 2006, 368, 1696.
- 2. Ahrén, B. Expert Opin. Emerging Drugs 2008, 13, 593.
- 3. Pei, Z. Curr. Opin. Drug Discov. Dev. 2008, 11, 512.
- 4. Gwaltney, S. L., II Curr. Top. Med. Chem. 2008, 8, 1545.
- Kim, D.; Wang, L.; Beconi, M.; Eiermann, G. J.; Fisher, M. H.; He, H.; Hickey, G. J.; Kowalchick, J. E.; Leiting, B.; Lyons, K.; Marsilio, F.; McCann, M. E.; Patel, R. A.; Petrov, A.; Scapin, G.; Patel, S. B.; Roy, R. S.; Wu, J. K.; Wyvratt, M. J.; Zhang, B. B.; Zhu, L.; Thornberry, N. A.; Weber, A. E. J. Med. Chem. 2005, 48, 141.
- Villhauer, E. B.; Brinkman, J. A.; Naderi, G. B.; Burkey, B. F.; Dunning, B. E.; Prasad, K.; Mangold, B. L.; Russell, M. E.; Hughes, T. E. J. Med. Chem. 2003, 46, 2774.
- Augeri, D. J.; Robl, J. A.; Betebenner, D. A.; Magnin, D. R.; Khanna, A.; Robertson, J. G.; Wang, A.; Simpkins, L. M.; Taunk, P.; Huang, Q.; Han, S.-P.; Abboa-Offei, B.; Cap, M.; Xin, L.; Tao, L.; Tozzo, E.; Welzel, G. E.; Egan, D. M.; Marcinkeviciene, J.; Chang, S. Y.; Biller, S. A.; Kirby, M. S.; Parker, R. A.; Hamann, L. G. J. Med. Chem. 2005. 48. 5025.
- Lübbers, T.; Böhringer, M.; Gobbi, L.; Hennig, M.; Hunziker, D.; Kuhn, B.; Löffler, B.; Mattei, P.; Narquizian, R.; Peters, J.-U.; Ruff, Y.; Wessel, H. P.; Wyss, P. Bioorg. Med. Chem. Lett. 2007, 17, 2966.
- Peters, J.-U.; Weber, S.; Kritter, S.; Weiss, P.; Wallier, A.; Boehringer, M.; Hennig, M.; Kuhn, B.; Loeffler, B.-M. Bioorg. Med. Chem. Lett. 2004, 14, 1491.
- Boehringer, M.; Kuhn, B.; Luebbers, T.; Mattei, P.; Narquizian, R.; Wessel, H. P. U.S. Pat. Appl. 2004/0259902, 2004.
- 11. Beke, D.; Szantay, C. *Chem. Ber.* **1962**, 95, 2132.
- 12. Kawatsura, M.; Hartwig, J. F. J. Am. Chem. Soc. 1999, 121, 1473.
- 13. Kuhn, B.; Hennig, M.; Mattei, P. *Curr. Top. Med. Chem.* **2007**, 7, 609.
- 14. Bendels, S.; Kansy, M.; Wagner, B.; Huwyler, J. Eur. J. Med. Chem. 2008, 43, 1581.
- 15. Lüllmann-Rauch, R.; Pods, R.; von Witzendorff, B. Toxicology 1996, 110, 27.
- 16. Hunziker, D.; Hennig, M.; Peters, J.-U. Curr. Top. Med. Chem. **2005**, 5, 1623.
- 17. The co-ordinates of the X-ray complex structure of human DPP-IV with compound (S,S,S)-2g have been deposited at the Protein Data Bank, accession code 3kwj.
- 18. Fischer, H.; Kansy, M.; Bur, D. *Chimia* **2000**, *54*, 640.
- Fischer, H.; Kansy, M.; Potthast, M.; Csato, M. Rational Approaches to Drug Design, Proceedings of the European Symposium on Quantitative Structure– Activity Relationships, 13th, Düsseldorf, Germany, Aug. 27–Sept. 1, 2000 2001, 286.
- 20. Mortelmans, K.; Zeiger, E. Mutat. Res. 2000, 455, 29.
- 21. Fenech, M. Mutat. Res. 2000, 455, 81.